

约束应激对大鼠 5-HT_{1A} 和 5-HT_{2A} 受体表达的影响

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Effect of Movement Restrict on the Expression of 5-HT Receptor Subtypes in Rat Brain

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【Abstract】 Objective: To explore the 5-HT mechanism of stress. Methods: The method of polymerase chain reaction (PCR) was used to detect the presence of 5-HT receptor subtypes in the hippocampus, hypothalamic and midbrain of the rat after 30 days of movement restrict compared with normal cage control groups (CC). The specific oligonucleotide primers were synthesized based on the complementary DNA sequence for each of the rat 5-HT_{1A} and 5-HT_{2A} receptor subtypes. Results: The expression of 5-HT_{1A} receptor subtypes in each regions of the rat brain after movement restrict were significantly lower than that of the CC groups and the expression of 5-HT_{2A} receptor subtype were significantly higher ($P < 0.01—0.05$). Conclusions: 5-HT_{1A} and 5-HT_{2A} receptors may be involved in the stress of the rat and may have opposite effects with respect to certain functions.

【Key words】 Stress; Rat; 5-HT receptor; PCR

慢性应激如长期压抑、环境不适等易导致身心健康损害, 出现学习记忆、情绪行为等多方面的改变^[1,2], 目前对其发生机制的研究多集中在下丘脑—垂体—肾上腺(HPA)轴、自主神经系统和免疫系统等, 而对 5-羟色胺(5-HT)的研究较少。5-HT 是一种重要的中枢神经递质, 参与多种行为、情绪活动的调节, 迄今已发现了 5-HT 受体的 7 种类型 13 个亚型。近来的研究报道认为 5-HT_{1A} 和 5-HT_{2A} 可能与精神机能和学习记忆有关^[3,4]。为探讨慢性应激与 5-HT_{1A}、5-HT_{2A} 受体的关系, 我们采用 PCR 观察大鼠经 30 天慢性约束应激后不同脑区 5-HT_{1A}、5-HT_{2A} 受体 mRNA 的表达情况, 对应激的 5-HT 机制进行初步探索。

1 材料和方法

1.1 实验动物及分组

成年、雄性、健康上海产 Sprague-Dawley 大鼠, 体重 200~250g (由第四军医大学实验动物研究中心提供), 随机分为约束应激组和正常笼养组, 每组 6 只, 共 12 只。实验前在研究室内分笼饲养 1 周, 食水自由。

1.2 实验方法

1.2.1 模型制备 根据文献^[5]制作应激模型。应激箱为 30cm×30cm×40 cm 大小的玻璃水箱, 箱中注满水, 正中立一直径 18.0 cm 圆形平台, 平台高出水面约 1.0 cm。大鼠只能在平台上活动, 可自由进

食进水和睡眠。对照组单独笼养。实验期间光照明暗周期 12 h/12 h, 室温(20±2)℃。箱中水温保持在 20℃左右, 每天换水以保持清洁。

1.2.2 组织准备 大鼠经 30 天约束慢性应激后予腹腔注射戊巴比妥钠(50 mg/kg)麻醉, 以焦碳酸二乙酯(DEPC)处理的 0.01 mol/L PBS 经心灌流冲洗血液, 快速取海马、下丘脑和中脑, 立即在干冰上冷冻, 保存于-80℃冰箱。

1.2.3 总 RNA 提取 采用异硫氰酸胍法提取总 RNA。首先将组织 1 ml TRI zol 试剂(GIBCO BRL)中用 Polytron 匀浆器(ART)匀浆, 室温孵育 5 min, 加入 0.2 ml 氯仿, 震荡混匀, 以 9000 r/min, 离心 10 min。取上层水相, 加等体积丙酮混合, 室温孵育 10 min。于 4℃以 15000 r/min 离心 15 min, 弃上清。沉淀块用 70%乙醇冲洗后, 以 20 μl DEPC 处理的去离子水溶解。将溶解的总 RNA 加入含 0.01 mol/L Tris-HCl、0.05 mol/L KCl、1.5 mmol/L MgCl₂、2.5 mmol/L LiDTT 和 1 μl DNase I(Boehringer Mannheim)的溶液中, 37℃水浴中孵育 1 h, 然后于 95℃处理 3 min, 用酚/氯仿/异戊醇抽提, 以乙酸钠/乙醇沉淀。将 RNA 沉淀溶解于 DEPC 处理水中, 并用分光光度计(Hitachi)测量 RNA 浓度, 于-80℃保存备用。

1.2.4 反转录合成 cDNA 在 50 μl 的反应体积中加入 5 μg 总 RNA、5× 反应缓冲液 10 μl, 10 mmol/L dNTPs 5 μl, RNasin(WAKO, 40 μg/μl)0.5 μl, oligo(DT) 12—18(GIBCO BRL)0.25 μg, 反转录酶(M-MLV,

GIBCO BRL, 200 μ /L)4 μ L, 0.1 mol/L DTT0.5 μ L, 置37℃孵育1h, 然后在65℃加热5min, 储存于-30℃。

1.2.5 引物设计及合成 按照已报道的5-HT_{1A}和5-HT_{2A}受体的cDNA序列^[3,4]设计其相应的特异性引物, 并由Applcia Biosystems Model 394DNA合成仪(Pharmacia)合成。合成的引物用18%的变性聚丙烯酰胺凝胶电泳进行检测。各引物的序列分别为, 5-HT_{1A}: 5'-TCA CCT GCG ACC TGT TTA TC-3', 位置320~339bp, 5'-GCT CCC TTC TTT TCC ACC TT-3', 位置694~713bp, 扩增产物长度394bp; 5-HT_{2A}: 5'-GCT CTT TTC TAC GGC ATC CAT C-3'; 位置468~489bp, 5'-AGT TCT TTT TCT GTC CCA CCT G-3', 位置1273~1294bp, 扩增产物长度827bp。

1.2.6 PCR扩增 在25 μ l的反应体积中加入合成的cDNA 0.1 μ g, 10 \times PCR缓冲液2.5 μ l, dNTPs(2mmol/L)2.5 μ l, MgCl₂(25 mmol/l)2.5 μ l, 各引物10 μ l(20 pmol/ μ l), TapDNA聚合酶(Toyobo, 5u/ μ l)0.

5 μ l。使用PTC-100PCR仪(MJ Research)进行热循环。循环条件: 93℃1 min, 56℃30 s, 72℃1 min, 进行1个循环; 93℃30 s, 56℃30 s, 72℃1 min, 进行35个循环; 72℃延伸8 min。取各扩增产物10 μ l, 经3%琼脂糖(FMC)凝胶电泳, 溴化乙锭染色后, 用UVP凝胶成像系统进行检测, 并用Labworks软件对各阳性条带的密度进行测定。以 β -actin为模板作为阳性内参照, 目的基因与 β -actin共同扩增。

2 结 果

以大鼠TGcDNA为模板的5-HT_{1A}、5-HT_{2A}受体亚型在约束应激大鼠及对照组大鼠海马、下丘脑和中脑的PCR扩增产物的琼脂糖胶电泳均呈现阳性条带, 两阳性条带的位置分别与预期的各亚型PCR产物相符。利用Labworks软件所做的分析表明, 应激组各脑区5-HT_{1A}受体mRNA表达显著低于对照组($P < 0.01$), 而5-HT_{2A}受体mRNA表达显著高于对照组($P < 0.01 \sim 0.05$), 见附表。

附表 应激组与对照组各脑区5-HT_{1A}、5-HT_{2A}mRNA相对水平比较(% β -actin)

组别	海马		下丘脑		中脑	
	5-HT _{1A}	5-HT _{2A}	5-HT _{1A}	5-HT _{2A}	5-HT _{1A}	5-HT _{2A}
正常对照组	46.7±6.7	65.7±5.8	58.6±7.5	62.9±7.5	52.5±6.8	54.2±6.8
约束应激组	73.6±8.4	43.7±5.5	76.8±9.2	35.4±5.6	83.7±8.9	30.2±4.3
t值	6.132	6.742	3.756	7.197	6.823	7.307
P值	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01

3 讨 论

5-HT是脑内一种重要的神经递质。释放5-HT的神经细胞仅位于脑干中缝的几组核团, 但它们发出的投射纤维则分布于大脑皮质、皮质下区、小脑、脑干及脊髓等广大部位的脑区。同样, 5-HT受体也广泛分布于整个中枢神经系统。我们采用PCR观察大鼠约束应激后不同脑区5-HT_{1A}、5-HT_{2A}受体mRNA的表达情况。结果显示, 两组大鼠海马、下丘脑和中脑存在5-HT_{1A}、5-HT_{2A}受体mRNA的表达。对5-HT_{1A}、5-HT_{2A}受体mRNA表达相对密度水平分析发现, 应激组各脑区5-HT_{1A}受体mRNA表达显著低于对照组, 而5-HT_{2A}受体mRNA表达显著高于对照组。与Lopez^[6]等报道应激后海马5-HT_{1A}受体mRNA的表达减少, 额叶HT_{2A}受体mRNA的表

达增加基本一致。

不同亚型的5-HT作用功能及调节机制不尽相同。研究发现5-HT_{1A}和5-HT_{2A}受体在调节情绪、认知、睡眠觉醒等方面的作用相反^[7], 对大脑衍生的神经生长因子(BDNF)的调节作用也不同^[8], 而BDNF对海马和其它大脑区域有重要的营养作用。Vaidya等人报告, 预先给予5-HT_{2A}拮抗剂使BDNF下调减少50%, 而5-HT_{1A}不具有该效应。另外的研究却发现, 应激可引起海马内5-HT_{1A}受体结合减少, 而这又与海马CA3区的萎缩和记忆损害有关。因此, 大鼠应激时齿状回和海马内BDNF mRNA的表达水平显著减少可能与应激后5-HT_{1A}生成减少和5-HT_{2A}增多有一定关系。

糖皮质激素可调节5-HT_{1A}和5-HT_{2A}受体且有不同的影响。给健康志愿者急性服用氢化可的松,

可显著降低 5-HT_{1A} 受体的功能，服用地塞米松则可引起与用量有关的 5-HT_{2A} 受体表达的增加^[9]。应激升高皮质酮水平可降低海马以下区域中 5-HT_{1A} 受体结合水平^[10]，而切除肾上腺则可增加海马的 5-HT_{1A} 受体密度^[11]。外源性的 ACTH 可提高新皮层皮质 5-HT_{2A} 受体的水平，肾上腺切除术可消除对 5-HT_{2A} 受体的影响。由此推测应激后脑内 5-HT_{1A} 和 5-HT_{2A} 受体 mRNA 的不同表达可能与下丘脑—垂体—肾上腺皮质(HPA)轴被激活有关。

综合本实验发现，慢性应激时海马、下丘脑和中脑 5-HT_{1A} 和 5-HT_{2A} 受体 mRNA 的表达发生变化，表明海马、下丘脑和中脑均参与了慢性应激的调节。但 5-HT_{1A} 和 5-HT_{2A} 受体 mRNA 的变化并不一致，提示 5-HT_{1A} 和 5-HT_{2A} 受体在应激反应中具有不同的功能，其作用机制有待进一步探讨。

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